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## Simple Transfection Reagent (Cat: T3080)

**Sizes:** 500uL (5x 100ul)

*Research use only, not for diagnostic use.*

**Storage:** -20°C or -80°C. • After thawing the solution can be stored at 4 °C for up to 2 months.

### Description:

Simple transfection reagent is a ready-to-use polymer-based transfection agent, with high transfection efficiency and lowest toxicity to deliver DNA or siRNA into various primary/ mammalian cell lines.

### Workflow Timeline:

Day 1: Seed cells

Day 2:(am/pm) Transfect Cells (am), After 2h-4h transfection - add the complete media (pm).

Day 3:(pm) or more Observe fluorescence, or by FACS analysis.

### General Protocol:

**Day 1: Cell Seeding:** Split cells one day before transfection.

a. **6 well plate:**  $2 \times 10^5$  cells/well

b. **10 cm dish:**  $1 \times 10^6$  cells/dish

The cell density should reach to the 40-90% confluency at the time of transfection.

**Day 2: (Am)** Prior to transfection, bring Simple-Transfection reagent (T3080) to room temperature and warm the serum-free media (DMEM w/o phenol red) or PBS to 37 °C.

#### 1. Preparation the complex: **Transfection agent (ul) + DNA(ug) (3:1).**

\* Dilute total plasmid DNA (ug)/siRNA in serum-free media (DMEM w/o phenol red) or PBS.

\* Add Simple Transfection agent to the diluted DNA/RNA. (3:1 ratio of T3050 (ul): DNA (ug))

a. For **6 well plate:** 300ul serum-free media (DMEM)+ **3 ug of DNA**, vortex briefly to mix well, add **9 ul of Simple- Transfection reagent;**

b. For **10cm dish:** 900uL serum-free media (DMEM)+ **9 ug of DNA**, vortex briefly to mix well, add **27ul of Simple-Transfection reagent;**

**2. Immediately pulse vortex for 10 seconds. Spin down briefly.**

**3. Incubate 15 minutes at Room Temperature (RT) for forming the TransFection/DNA or RNA complexes.**(The formation of TransFection/DNA might make solution slightly cloudy).

**4. Aspirate the cell culture medium from each well/dish and add the Transfection/DNA complexes to cells carefully drop-wise.** Gently rock the plate to ensure even distribution.

Do not swirl plate and not to dislodge the cells.

**5. Return the well/dish to a 37 °C incubator with 5% CO<sub>2</sub>.**

**Day 2: (Pm) Incubate at 37°C for 2h–4h**, add appropriate volumes of complete cell culture medium to the well/ dish and continue to incubate the cells for 12-24 h.

a. For **6 well plate:** add 2 mL complete medium/each well.

b. For **10 cm dish:** add 9 mL complete medium/dish.

**Day 3: (Pm) or more:** Observe fluorescence, harvest cells, or perform your experiment by FACS analysis.

**Precautions and Disclaimer:** This product and procedure described are intended for R&D use only. Purchase of this product does not convey a license to perform any patented process.